Complex strategies with Genomic Colocation Exercise 14

14.1 Divergent genes with similar expression profiles. Note: for this exercise use http://plasmodb.org.

Identify genes that meet these four criteria:

- 1. are located within 1000 bp of each other
- 2. are divergently transcribed,
- 3. are expressed maximally at day 30 of the iRBC cycle +- 8 hrs and,
- 4. show at least a 3-fold increase in expression.
- Hint: first use the "Genes bases on Microarray Evidence" -> "Erythrocytic expression time series (3D7,DD2, & HB3) (Bozdech et al. and Linas et al.)" -> "Fc" search.

Identify Genes based on P.f. Intracrythroc For the Experiment iRBC HB3 (48 Hour scaled) ÷ return protein coding ÷ Genes that are up-regulated ÷ with a Fold change >= 3	Example showing one gene that would meet search criteria (Dots represent this gene's expression values for selected samples) Up-regulated
between each gene's minimum : expression value? in the following Reference Samples ? select all clear all expand all collapse all reset to default : 17-30 Hours : 17-23 Hours : 24-30 Hours : 31-48 Hours : and its maximum : expression value? in the following Comparison Samples ? : select all clear all expand all collapse all reset to default : 17-30 Hours : eslect all clear all expand all collapse all reset to default : 17-30 Hours : 17-30 Hours	Image: the search calculates:Image: th
🗆 Advan	ced Parameters
Weight ? 10 Global min / max in selected time points ? Maximum	•
Get	Answer

 Add a step that is the same as the first step and select the genomic colocation (1 relative to 2) operation.

- Set up the form to identify those genes that are transcribed on the opposite strand that have their starts located within 1000 bp of another genes start.
- If you are having difficulty setting this up, you can see the strategy at: <u>http://plasmodb.org/plasmo/im.do?s=0ebfe58b1c9b42cc</u>. Cut and paste the link into your browser if the hyperlink does not work.
- Turn on the "Pf-iRBC 48hr Graph" column to assess how well the pairs of genes compare in terms of expression. The pairs of genes are located one above the other in the result table if sorted by location.
- Note that you could do similar types of experiments to look at potential coregulation / shared enhancers / divergent promoters with other sorts of data such as:
 - Genes by ChiP-chip peaks in ToxoDB.
 - DNA motifs for transcription factor binding sites.
 - Of course other expression queries.
 - o Etc ...
- The screenshot below shows one way (there are MANY) to configure the genome colocation form to identify genes that are divergently transcribed located with their start within 1000 bp of each other.

(FID)		ntive locations in the genome or Genes search (Step 2) returned 684 Genes.	TUTORIALS
(684 Genes in Step)	overlaps 🛟 the	upstream region of a Gene in Step 2 and is on the opp	posite strand 🛟 "
Region Gene	II	Region Gene	
 Exact Upstream: 1000 bp Downstream: 1000 bp 		 ○ Exact ⊙ Upstream: 1 bp ○ Downstream: 1000 bp 	
Custom: begin at: <u>start ♀</u> - ♀ 1000 bp end at: <u>start ♀</u> - ♀ 1 bp		Custom: begin at: start ↓ - ↓ 1 bp end at: start ↓ - ↓ 1 bp	
	Submit		Close

14.2 Finding possible oocyst expressed genes based on DNA motifs.

Note: for this exercise use <u>http://toxodb.org</u>

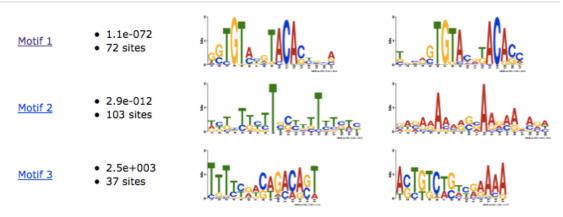
In exercise 13.4 you defined a number of *T. gondii* genes that are preferentially expressed in the oocyst stages. How can you use this information to expand the number of possible oocyst regulated genes? One possibility is to try and define

common elements in promoter or 5'UTR regions (ie. 5' to the start of the genes). For this you will have to be able to retrieve 5' sequence from all of the genes in the oocyst list. How would you do this? (hint: click on download genes then select FASTA format from the drop down menu). The amount of upstream sequence you retrieve is up to you.

After you have your sequences you will need to run them through a DNA pattern finder like MEME (<u>http://meme.sdsc.edu/meme/intro.html</u>). Results from a submission to MEME could take up to several hours so for your convenience 300 nucleotides upstream of all the oocyst results were analyzed using MEME – results can be visualized here:

Can you take one of the generated motifs and find additional genes in *T. gondii* that contain this motif in their upstream regions? What do your results look like? Did you get too many or too few results? How would you modify the motif to change your results?

Motif Overview



14.3. Identifying conserved DNA elements upstream of genes

The goal of this exercise is to identify a DNA element in the upstream region of similarly regulated genes.

a. Identify genes that are up-regulated in malaria sporozoites compared to blood stage parasites. Examine the list of searchable experiments on the PlasmoDB microarray search page: Identify Genes based on Microarray Evidence. Can you identify an experiment that would give you this answer? (hint: look at *Plasmodium* species other than *P. falciparum*, ie. *P. yoelii* [Liver, mosquito and blood stage expression profiles (Tarun et al.) (direct comparison)]

 Organism 	≎ Data Set		Choose a search
P. yoelii yoelii 17X	Liver, mosquito and blood s	Do	
		± Show All Data Sets ±	
Direct Comparison Pe	ercentile		
	Identify Genes	based on P.y. Liver Stages	(fold change)
	Direction 😯	up-regulated +	
	Samples 🕖	sgSpz vs BS +	
	Fold difference >= 😢	4	
	Protein Coding Only: 🕖	protein coding 💠	
		Advanced Parameters	
		Get Answer	

b. How many genes did you find? What you are interested in is looking at the nucleotide sequence upstream of the start sites of these genes. How can you do this in bulk? PlasmoDB has a sequence retrieval tool that allows you to download results of your searches in bulk. This includes a tool that allows you to specify the sequence you want.

(Genes)													Strategy: Py Expression(3) * 🛛 🔀
Rename Delete Save As Step 1 Add Step														
								-	_					
Strat	egy: Py	om Step Express	ion(3)	limit/filter yo									А	Add 57 Genes to Basket Download 57 Genes
				innumiter yo	urresults		Plasmodiur	n						1
All	Ortholog	P.berghei	P.chabaudi	P.cvnomolai	P.falciparum	(nr Genes: 0)	P.gallinaceum	P.knowlesi	P.reichenowi	P.vivax	vax P.yoelii (nr Genes: 57)		-	
Results	Groups	ANKA	chabaudi	strain B	3D7	п	8A	strain H	Dennis	Sal-1	yoelii 17XNL	yoelii 17X		
57	57	0	0	0	0	0	0	0	0	0	0	57	0	
[Results	Genome xt Last		Advanced Pagi	ing									Add Columns
	Gene I	D	Produc	t Descriptio	on 🎱 🔒					\$	Fold Change	0	Py-Live	er Stages - Graph 🕹
÷	PY17X_052	3600	conserved F	Plasmodium pr	rotein, unknow	n function					34.55		Extraction Velue (log2 rate	

 c. After you click on "Download ### Genes", you are offered a drop down menu of options. Explore these; which one will allow you to specify the sequence to download. (hint: Configurable FASTA)

Download 75 Genes from the search:				
P.y. Liver Stages (fold change)				
Please select a format from the dropdown list te **Note: Gene IDs will automatically be included in ✓ Select a format Tab delimited (Excel): choose from columns Text: choose from columns and/or tables Configurable FASTA GFF3: Gene models and optional sequences	create the download report. the report. EuPathDB	Please Contact Us with any questions or comments		
XML: choose from columns and/or tables json: choose from columns and/or tables	d? 🔘 A < 🌞 🦃 🛞 🍬 👌	Strategies WDK		

d. Define the sequence you want to retrieve. For this exercise retrieve 500 nucleotides up-stream of the start of translation.

Dow	nload 75 Genes from the search:				
	P.y. Liver Stages (fold change)				
Please select a format from the dropdown list to create the download report. **Note: Gene IDs will automatically be included in the report.					
Configurable FASTA \$					
This reporter will retrieve the sequences of the genes in your result. Choose the type of sequence: <a>genomic protein CDS <a>transcript Choose the region of the sequence(s):					
begin atTranslation Start (ATG)++500nucleotidesend atTranslation Start (ATG)++0nucleotides					
Download Type: Osave to File Oshow in Browser					
*** Note: If UTRs have not been annotated for a gene, then choosing "transcri	Get Sequences ption start" may have the same effect as choosing "translation start".				

e. The next step is to take this sequence and run it through a DNA motif finder such as MEME (<u>http://meme.sdsc.edu/meme/intro.html</u>). To speed up this process we have pre-run the motif finder and results are presented here:

Motif Overvio	ew		
<u>Motif 1</u>	5.5e-05846 sites		¹ 4884880 A8848448444
<u>Motif 2</u>	 2.8e-029 46 sites		A ATATAA A ATAAA ATAAA
<u>Motif 3</u>	9.3e-0055 sites	* Stan arg t <mark>own tattattatta angalast tool craac</mark>	* C TCIC A CAMATALS ATA MATANA TA LA LA GOCAL ALC

The regular expression for each of these motifs is presented here:

Motif 1:

TTT[TAG]T[TA]T[CT][TA][TC][TC][ATC]TTTTT[TG]TTT[TC][TA]TTT[TA]TTTT[TA]T[T C][TA][TC][TA][TC]TT[TC]

Motif 2:

[TC]A[TC][AT][TC]AT[ATG]T[GTA][TC][AG][TA][GAT][TC][GA]T[AGT]T[GA][TC]AT[AG]T[GAT][TC][AT]T

Motif 3:

[GAC][AG][TC]AT[AG][TC][GA]T[TG][GT][TCG]CCA[TG][AG]A[TG][AG]A[TA][TG][T A][AT][TG][TG][AC]T[AGT][TC]A[CAT][AG][TA][AT][ACG][TCG]T[TA][CA]A[TC][GA CTA][GC][TG][GA][AG]A[GC]

f. Can you find any of these motifs in the *P. yoelii* genome? (hint: use the DNA motif query)

dentify Other Data Types:	entify Genomic Segments based on DNA Motif Pattern
Isolates Isolates Genomic Sequences Genomic Segments (DNA Motif) DNA Motif Pattern Genomic Location P.f. eQTL HB3-Dd2 cross (segments by association to genes) SNPs ESTs ESTs ORFs	ganism V select al char al expand al collapse all reset to default Plasmodium berghel Plasmodium thabaudi Plasmodium falciparum Plasmodium gallinaceum Plasmodium reichenowi Plasmodium reichenowi Plasmodium vivax R M Plasmodium vivat R M Plasmodium vielai select al char al expand al collapse all reset to default Pattern CTITICA/(TC)AT(AGTTCAT)(TC)(AT)(TC)
H SAGE Tags	Give this search a weight
	Give this search a name
	Get Answer

g. How many times did this motif occur in the genome? How many of them are in the upstream region of genes? Can you find all *P. yoelii* genes that are within 1000 nucleotides downstream of the motif? (hint: use the genomic colocation option when combining searches).

	G	enomic Coloc	ation 🕄 🙄							
	Combine Step 1 and Step 2 using relative locations in the genome									
	You had 1257 Genomic Segments in your Strategy (Step 1). Your new Genes search (Step 2) returned 7774 Genes.									
"Return each Ge	Gene from Step 2 + whose upstream region	overlaps + t	he exact region	of a Genomic Segment in Step 1 and is or	either strand 🗘 "					
	(7774 Genes in Step)		·	(1257 Genomic Segments in Step)						
H	Region	<u> </u>		Region						
	Gene			Genomic Segment						
OE	Exact		 Exact 							
ΘU	Upstream: 1000 bp		OUpstream: 1	1000 bp						
OD	Downstream: 1000 bp		Oownstream	1000 bp						
Oc	Custom:		OCustom:							
1	begin at: start +) - +) 1000 bp end at: start +) - +) 1 bp		begin at: (end at: (
		Submit			Class					

h. Do these genes have orthologs in other *Plasmodium* species? (hint: add a step to your search strategy and transform the results to their orthologs).

	Add Step	×
Run a new Search for Transform by Orthology Add contents of Basket Add existing Strategy Filter by assigned Weight	Add Step 4 : Transform by Orthology	
	Population Biology	Close

Optional: add a step and do the motif search on these orthologs to find out how many of them also contain the motif.