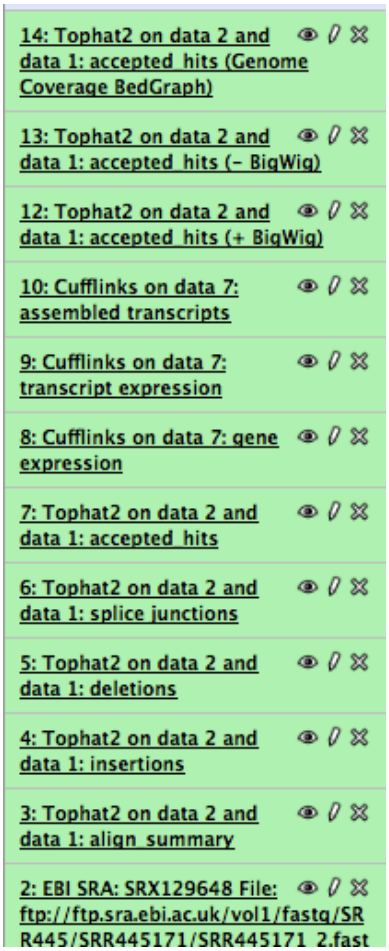
# Interpreting RNAseq Mapping results
# (Part 2: Loading data generated by the pathogen portal's RNAseq pipeline in the Genome Browser)
# Exercise 11

For this exercise we will be using:
http://pathogenportal.org
http://plasmodb.org




1. Explore the results of the RNA-sequence pipeline. What files were generated? To view contents of any of the results, click on the eye icon (  ) next to the file name.

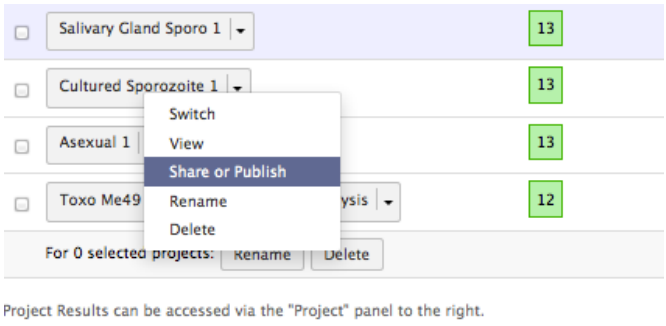
!!! important note – do not click on the icon next to the file called "Tophat2 on data 1 and data 3: accepted_hits" – this file is huge and will not display but rather will download the contents to your computer.

a. TopHat in RNArocket generates five files:
   *Align_summary*: this includes a summary of how the alignment went (ie. the number of reads that were aligned).
b. *Insertions*: reported insertions.
c. *Deletions:* reported deletions.
d. *Splice junctions*: reported junctions. Each junction consists of two connected BED blocks, where each block is as long as the maximal overhang of any read spanning the junction. The score is the number of alignments spanning the junction.
e. *Accepted hits*: BAM file (binary alignment map). Note that many alignment programs will generate a file called a SAM file (sequence alignment map) which is a table including text of the alignment and mapping. However, for viewing results in a sequence browser like GBrowse, the file needs to be converted into the binary formatted (BAM) – you do not have to worry about this for this exercise.

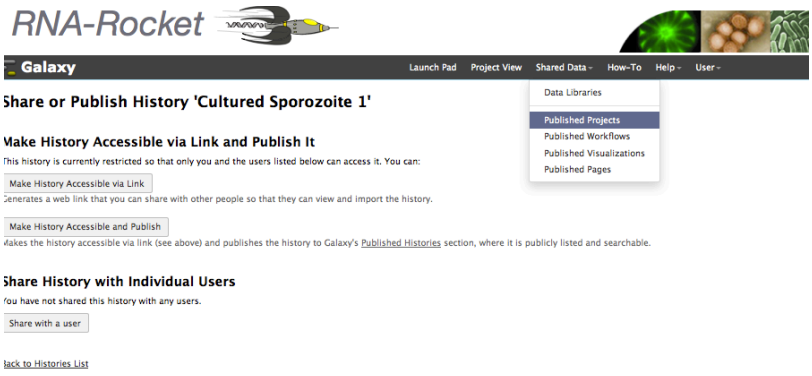
Cufflinks generates three files:
gene expression, transcript expression and assembled transcripts. The gene

expression and transcript expression files for our purposes should be identical since EuPathDB genomes do not have separate genes and transcripts. These files include the FPKM values (Fragments Per Kilobase of transcript per Million mapped reads) for each gene in the genome analyzed – in this case *Giardia* assemblages.
Additional files include files of the format BigWig and BedGraph. You can read more about these file formats here:
http://genome.ucsc.edu/goldenPath/help/bigWig.html
In a nutshell, these are file formats created from large binary files like BAM files and makes it possible to load these data in a genome browser.

**Note:** *to share your data with the rest of the workshop, select "Share or Publish" from the drop down menu on project you want to share. On the next page click on "Make History Accessible and Publish".*




*To import a history, select Published Projects from the Shared data menu item. Select the project you want to import then click on import history in the upper right hand side of the screen.*

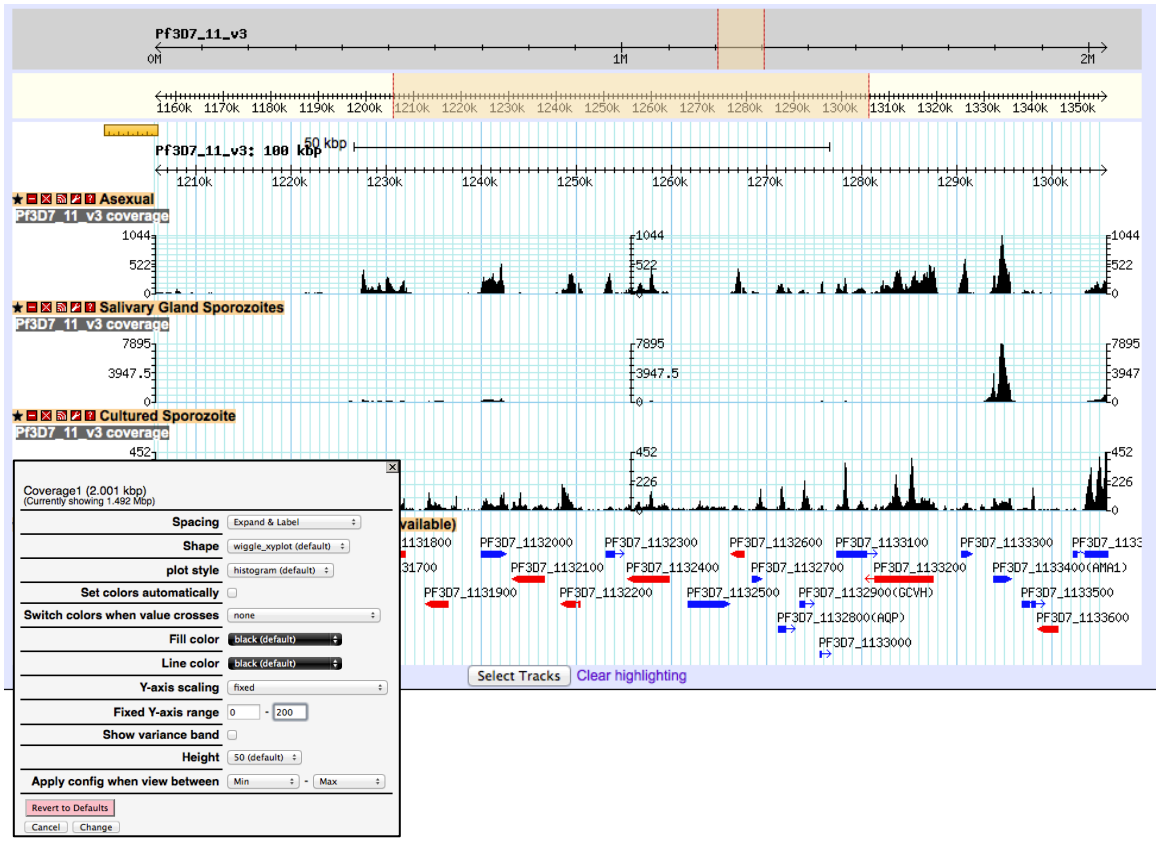


2. Load your BAM data (accepted hits) into GBrowse. Click on your “Tophat2 on data 2 and data 1: accepted_hits” in your project history panel. This will show you information about the file including a link to display data in GiardiaDB – click on the link.

3. Load the assembled transcript data. Essentially use a similar procedure as above.

4. Wait a couple of minutes for GBrowse to load your data.

5. Once data has been loaded, you can configure the track display settings. For example, you can adjust the Y-axis scaling to a fixed axis.


Pf3D7_11_v3
0M
1M
2M
1160k 1170k 1180k 1190k 1200k 1210k 1220k 1230k 1240k 1250k 1260k 1270k 1280k 1290k 1300k 1310k 1320k 1330k 1340k 1350k
Pf3D7_11_v3: 100 kbp
50 kbp
1210k 1220k 1230k 1240k 1250k 1260k 1270k 1280k 1290k 1300k
Asexual
Pf3D7_11_v3 coverage
1044
522
0
Salivary Gland Sporozoites
Pf3D7_11_v3 coverage
7895
3947.5
0
Cultured Sporozoite
Pf3D7_11_v3 coverage
452
226
0
Coverage1 (2.001 kbp)
(Currently showing 1.492 Mbp)
Spacing
Expand & Label
Shape
wiggle_xyplot (default)
plot style
histogram (default)
Set colors automatically
Switch colors when value crosses
none
Fill color
black (default)
Line color
black (default)
Y-axis scaling
fixed
Fixed Y-axis range
0
200
Show variance band
Height
50 (default)
Apply config when view between
Min
Max
Revert to Defaults
Cancel
Change
vailable)
1131800
PF3D7_1132000
PF3D7_1132300
PF3D7_1132600
PF3D7_1133100
PF3D7_1133300
31700
PF3D7_1132100
PF3D7_1132400
PF3D7_1132700
PF3D7_1133200
PF3D7_1133400(AMA1)
PF3D7_1131900
PF3D7_1132200
PF3D7_1132500
PF3D7_1132900(GCVH)
PF3D7_1133500
PF3D7_1132800(AQP)
PF3D7_1133600
PF3D7_1133000
Select Tracks
Clear highlighting