

RNA sequence data analysis


(Part 2: Loading data generated by the pathogen portal's RNAseq pipeline in the Genome Browser)

Exercise 10

For this exercise we will be using:

<http://pathogenportal.org>

<http://microsporidiadb.org>

1. Explore the results of the RNA-sequence pipeline. What files were generated? To view contents of any of the results, click on the eye icon () next to the file name.

!!! important note – do not click on the icon next to the file called “Tophat2 on data 1 and data 3: accepted_hits” – this file is huge and will not display but rather will download the contents to your computer.

TopHat generates four files:




























insertions, deletions, splice junctions and accepted hits. The accepted hits file is the BAM file (binary alignment map).

Note that many alignment programs will generate a file called a SAM file (sequence alignment map) which is a table including text of the alignment and mapping. However, for viewing results in a sequence browser like GBrowse, the file needs to be converted into the binary formatted (BAM) – you do not have to worry about this for this exercise.

Cufflinks generates three files:

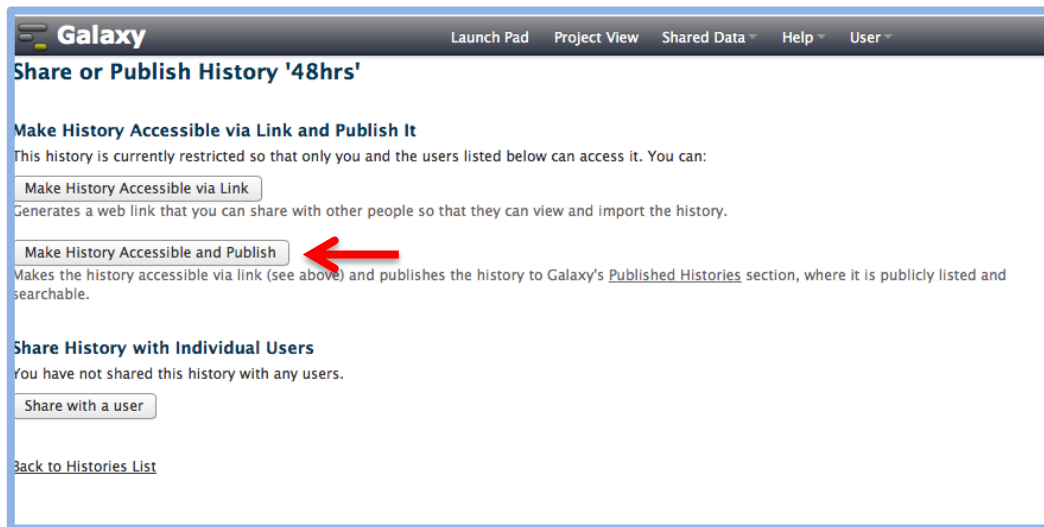
gene expression, transcript expression and assembled transcripts. The gene expression and transcript expression files for our purposes should be identical since EuPathDB genomes do not have separate genes and transcripts. These files include the FPKM values for each gene in the genome analyzed – in this case *Encephalitozoon cuniculi* ECII.

2. Share your accepted hits files. Click on the drop down menu for your project and select the option “share or publish”.

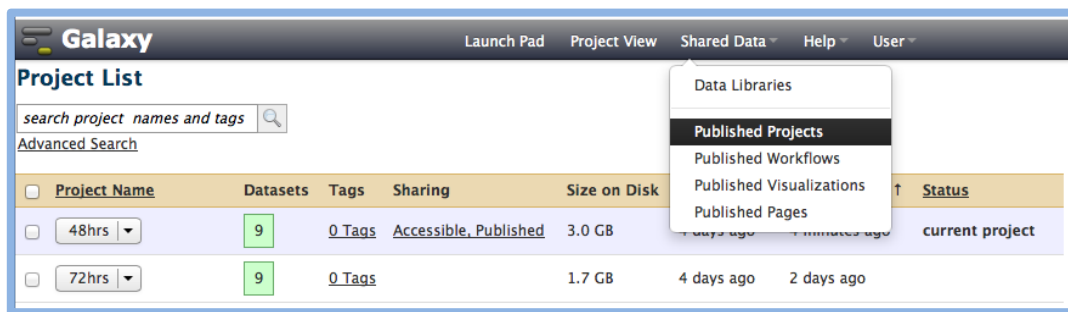
10: Cufflinks Eukaryotic on data 7: assembled transcripts			
9: Cufflinks Eukaryotic on data 7: transcript expression			
8: Cufflinks Eukaryotic on data 7: gene expression			
7: Tophat2 on data 1 and data 3: accepted hits			
6: Tophat2 on data 1 and data 3: splice junctions			
5: Tophat2 on data 1 and data 3: deletions			
4: Tophat2 on data 1 and data 3: insertions			
3: ftp://ftp.ddbj.nig.ac.jp/ddbj_databases/dra/fastq/SRA061/SRA061150/SRX229331/SRR769606_1.fastq			
1: ftp://ftp.ddbj.nig.ac.jp/ddbj_databases/dra/fastq/SRA061/SRA061150/SRX229331/SRR769606_2.fastq			

9	0 Tags	3.0 GB	4 days ago	4 days ago	current project
2	0 Tags	0 bytes	4 days ago	4 days ago	
8	0 Tags	0 bytes	6 days ago	6 days ago	
Babesia_bovis	0 Tags	Accessible, Published	853.0 MB	Jun 15, 2012	May 12, 2013
For 0 selected projects: Rename Delete					

On the next page select the option: Make History Accessible and Publish



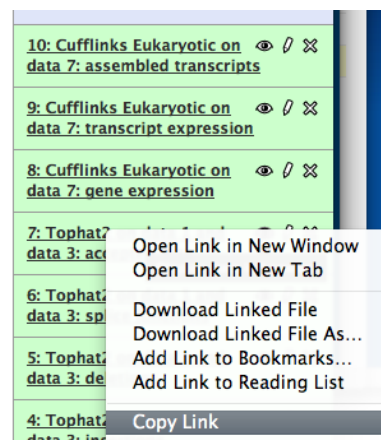
Once your project is published other people can access it by going to “Published Projects” section under the Shared data menu option in the Galaxy menu bar.



3. Load your BAM data into GBrowse. Navigate to the genome browser in MicrosporidiaDB and choose a landmark for *Encephalitozoon cuniculi* ECII you can just cut and paste the following into the “landmark or region” box:
ECII_CH11:98,571..148,570

Next, do the following to copy the link to the tophat accepted hits in pathogenportal to GBrowse:

- Control click (same as right click on a windows machine) on the eye icon for the tophat accepted hits.
- In GBrowse click on the “Custom Tracks” tab.



Browser | **Select Tracks** | **Snapshots** | **Custom Tracks** | **Preferences**

Search

Landmark or Region :

Examples : AL590443:85000-115000, ECI_CH11:115000..135000.

- c. Click on the “From a URL” link and paste the link you copied from pathogenportal.

Browser | **Select Tracks** | **Snapshots** | **Custom Tracks** | **Preferences**

Custom Tracks

[\[Help with uploading custom tracks\]](#)
 There are no tracks yet.
 Add custom tracks : [\[From text\]](#) [\[From a URL\]](#) [\[From a file\]](#)

Fetch track file from this URL

- d. Delete the last portion of the URL: display/?preview=True

Browser | **Select Tracks** | **Snapshots** | **Custom Tracks** | **Preferences**

Custom Tracks

[\[Help with uploading custom tracks\]](#)
 There are no tracks yet.
 Add custom tracks : [\[From text\]](#) [\[From a URL\]](#) [\[From a file\]](#)

Fetch track file from this URL

- e. Click on import.....and be patient.

Browser | **Select Tracks** | **Snapshots** | **Custom Tracks** | **Preferences**

Custom Tracks

[\[Help with uploading custom tracks\]](#)
http_rnaseq.pathogenportal.org_datasets_...
[\[http_rnaseq.pathogenportal.org_datasets_b8b5c2c15db111b6_\]](#) Imported
 Click to add a description

Source files:			
http_rnaseq.pathogenportal.org_datasets_b8b5c2c15db111b6_	Tue Jun 4 01:06:27 2013	579480617 bytes	
Configuration	Tue Jun 4 01:09:06 2013	1425 bytes	[edit]

Add custom tracks : [\[From text\]](#) [\[From a URL\]](#) [\[From a file\]](#)

- f. Once the data has loaded click on the Browser tab to view your data.

4. Load the assembled transcript data. Cufflinks generates this file in a format called GFF. This format is not accepted by GBrowse so you have to convert it to another format called BED. To do this click on the pencil icon next to the file. Click on “Covert Format” then click on convert. A new file will be generated in BED format. You can not copy the link to the file and load it into GBrowse the same way you loaded the BAM file.

The screenshot illustrates the process of converting a GFF file to a BED file within the GBrowse interface. The workflow is as follows:

- Dataset List:** A list of datasets is shown on the left. Dataset 10, "Cufflinks Eukaryotic on data 7: assembled transcripts", is highlighted. It has a pencil icon for editing attributes.
- Edit Attributes Panel:** Clicking the pencil icon opens the "Edit Attributes" panel. It has tabs for "Attributes", "Convert Format", "Datatype", and "Permissions".
- Convert Format Panel:** The "Convert Format" tab is selected. It shows the "Convert to new format" section with a dropdown menu set to "Convert GFF to BED". Below this, a "Convert" button is visible.
- Resulting Dataset:** After clicking "Convert", a new dataset, "12: Convert GFF to BED on data 10", is added to the top of the dataset list.

Additional details from the interface include:

- Name:** Cufflinks Eukaryotic on data 7: assem
- Info:** cufflinks v2.0.2
cufflinks -q --no-update-check -l
- Annotation / Notes:** (Empty field)
- Auto-detect:** This will inspect the dataset and attempt to correct the above column values if they are not accurate.